

**QUALITY CONTROL OF TWO CINNAMON TEA BRANDS  
COMMERCIALIZED IN COSTA RICA USING THE CENTRAL  
AMERICAN TECHNICAL REGULATION 11.03.56:09**

**Ariana Araya-Mena<sup>1</sup>, David Arias-Nuñez<sup>1</sup>, Karla Coto-Arce<sup>1</sup>, María Rebeca López-Hidalgo<sup>1</sup>, Michelle Villalobos-Varela<sup>1</sup>, Arlene Loría-Gutiérrez<sup>2</sup>, Jeimy Blanco-Barrantes<sup>3,4</sup> and Juan José Mora-Román<sup>4\*</sup>**

<sup>1</sup>Pharmacy Student, Faculty of Pharmacy, Universidad de Costa Rica, San José, Costa Rica.

<sup>2</sup>Pharmacology, Toxicology, and Pharmacodependence Department, Faculty of Pharmacy, Universidad de Costa Rica, San José, Costa Rica.

<sup>3</sup>Laboratory of Pharmaceutical Analysis and Assessment (LAYAFA), Universidad de Costa Rica, San José, Costa Rica.

<sup>4</sup>Industrial Pharmacy Department, Faculty of Pharmacy, Universidad de Costa Rica, San José, Costa Rica.

Article Received on  
19 August 2020,

Revised on 09 Sept. 2020,  
Accepted on 29 Sept. 2020,

DOI: 10.20959/wjpr202012-18895

**\*Corresponding Author**

**Juan José Mora-Román**

Industrial Pharmacy  
Department, Faculty of  
Pharmacy, Universidad de  
Costa Rica, San José, Costa  
Rica.

**ABSTRACT**

A tisane is a pharmaceutical form used in herbal teas. One product commercialized in this way in Costa Rica is cinnamon. Therefore, this investigation sought to determine the quality of two cinnamon tea brands produced and commercialized in Costa Rica and determine reproducibility in terms of the desired quality characteristics between different brand batches. Said determination was done by performing tests (described in official books, according to Costa Rican regulations) on three batches of each brand: labeling, organoleptic, foreign matter, minimum fill, heavy metals limit (lead), arsenic limit, loss on drying, total ash, acid-insoluble ash, microbial enumeration and specific microorganisms (*E. coli* and *Salmonella* sp.). The evaluated batches

complied with the specifications for each one. The exception was the labeling test (two and nine requirements were absent in both brands' primary and secondary packaging, respectively). Besides, there was reproducibility in terms of quality specifications for the three batches of the same product.

**KEYWORDS:** natural product, tisane, cinnamon, quality control.

## INTRODUCTION

Throughout history, natural products have been an abundant source of biologically active compounds for the pharmaceutical industry.<sup>[1]</sup> According to fossil records, plants for treating diseases can be traced back to around 60,000 years.<sup>[2]</sup> Its value lies in the ability to influence various signaling pathways.<sup>[1]</sup>

An example is the tisane, the pharmaceutical form of herbal teas.<sup>[3]</sup> Typically, an infusion, where the substance of interest is extracted by contacting boiling water with the herbal material, is done. The contact time is five to 20 minutes.<sup>[4]</sup>

A product commercialized in this pharmaceutical form in Costa Rica is cinnamon (*Cinnamomum verum* or *Cinnamomum zeylanicum*). It is a perennial tree with numerous flowers, whose leaves have three to five veins, which go from the base and almost to the tip's end. In addition, they are highly variable in size and shape.<sup>[5] [6]</sup>

Among the main constituents of this species are cinnamaldehyde, eugenol, and camphor. They are in different proportions, depending on the part of the plant selected. The traditional indications include gastrointestinal and gynecological conditions. Other attributable properties are decreased blood glucose levels and blood pressure, anti-inflammatory, antibacterial, and antioxidant effects.<sup>[7]</sup>

Due to their origin, natural products are considered options that generate benign adverse effects,<sup>[1]</sup> so most of the population prefers their consumption. However, this reasoning is wrong, since the consideration that they do not generate toxicity and are free of adverse effects has caused an incorrect use in unrestricted doses, resulting in severe poisonings and acute health problems.<sup>[8]</sup>

All pharmaceutical products marketed, including those obtained through natural sources, must be safe and effective at their commercialization and useful life. A drug's safety refers to a favorable benefit-risk ratio, while efficacy is the degree of benefit that offers under ideal conditions of employment.<sup>[9]</sup> For this reason, quality control favors monitoring both its efficacy and its safety.<sup>[10]</sup> In the absence of such control, situations such as erroneous plant identification, contamination, plant substitution or adulteration, failures in Good Manufacturing Practices (GMP), and incorrect doses can occur.<sup>[11]</sup>

Given this scenario, this research's objective was to determine the quality of two cinnamon tea brands produced in Costa Rica and are currently commercialized in the local market. Previously, such determination was performed with other Costa Rican products, specifically valerian,<sup>[12]</sup> senna leaf,<sup>[13]</sup> mint,<sup>[14]</sup> and boldo.<sup>[15]</sup> Moreover, reproducibility was determined in terms of the desired quality characteristics between different brand batches. All the investigation follows the Central American Technical Regulation (RTCA, for its Spanish acronym) 03.11.56:09.<sup>[16]</sup>

## **MATERIALS AND METHODS**

### **Product sampling and procedures selection**

Three batches of cinnamon tisanes of two brands were purchased in different supermarkets of Costa Rica. The batches were identified as 1, 2, and 3 for brand 1 and 4, 5, and 6 for brand 2.

The official books established by the RTCA 11.03.56.09<sup>[16]</sup> were employed to select the following procedures.

### **Labeling test**

The items stipulated for the primary and the secondary packaging were examined. The labeling of each batch must comply with the RTCA 11.04.41:06.<sup>[17]</sup>

### **Organoleptic characteristics test**

The odor and the color were determined as established by the European Pharmacopeia 10.0.<sup>[18]</sup> Ten tisanes were observed with a Konus® luminous lens. The product complied by presenting a yellowish or reddish-brown color and a characteristic, aromatic odor.

### **Foreign matter determination test**

Ten tisanes were opened, and their contents dispersed on a white surface and visualized with the help of a Konus® luminous magnifying glass. The foreign matter was separated, its weight was obtained with the help of an Adam® PW254 analytical balance, and its percentage was calculated regarding the total sample weight. The product complied if the foreign elements did not exceed 2%.<sup>[5]</sup>

### **Minimum fill test**

It was performed according to the general chapter <755> of the United States Pharmacopeia (USP) 42.<sup>[19]</sup> Ten tisanes from each batch were taken and open carefully. Then, the specific content of each tisane was weighed in an Adam® PW254 analytical balance. The acceptance

specification is that any individual tisane's net content was not less than 90% of the declared amount.

#### **Lead limit test**

The evaluation of heavy metals was made for the lead element by following method A presented in the <2.4.8> chapter of the European Pharmacopoeia 10.0.<sup>[18]</sup>

Two tisanes were taken in a 100 ml beaker, and 22 ml of distilled water were added. It was left to rest for 5 minutes. After this, the liquid was extracted from both tisanes, depositing them in the same beaker. Later, a 12.0 ml aliquot was taken for the sample, 2.0 ml for the standard, and 2.0 ml for the blank. Each one was tip out in a different test tube. Then, 10 ml of the lead standard (1 ppm) and 10 ml of distilled water were added to the standard tube and the blank tube, respectively. Subsequently, 2 ml of acetic acid-sodium acetate buffer at a 3.5 pH, 0.5 ml of diluted acetic acid, and 1.2 ml of thioacetamide solution (previously heated in a steam bath) were added to each tube. The result complied if both the blank and the sample had a brown color of less intensity than the standard.

#### **Arsenic limit test**

The test was done according to method A of the <2.4.2> chapter of the European Pharmacopoeia 10.0.<sup>[18]</sup>

In a 100 ml beaker, one tisane was placed with 11 ml of distilled water and left to rest for 5 minutes. After this, the liquid was extracted by compressing the tisane and deposited in the same beaker. Later, a 10.0 ml aliquot was taken and transferred in a conical flask. Afterward, 15 ml of 12 M HCl, 0.1 ml of stannous chloride, and 5 ml of 16% w/v potassium iodide were tip out. It was left to stand for 15 minutes. Then, 3 g of zinc chips were added, and the apparatus was assembled and placed in a water bath (temperature range between 90 and 100 °C). The standard was prepared in the same way, utilizing 1 ml of standard arsenic solution (1 ppm). After not less than 2 hours, the stain produced on the sample's mercury bromide paper should not be more intense than the reference standard.

#### **Loss on drying test**

It was carried out following the general chapter <731> of the USP 42.<sup>[19]</sup> A crucible was dried for 30 minutes at 105 °C in a Thermo Scientific® HeraTherm oven stove. The process was repeated until constant weight. Next, a cinnamon tisane's content was placed in the

crucible and accurately weighed on an Adam® PW254 analytical balance. Then, it was placed in the oven and dried for two hours at 105 °C. After cooling in a desiccator, it was weighed again in the same balance to calculate the loss percentage. These periods of heating and cooling were continued until reaching a constant weight. The product complied if the loss on drying percentage was no more than 12% from the initial weight.

#### **Total ash test**

The procedure of the general chapter <561> of the USP 42 was utilized.<sup>[19]</sup> Two cinnamon tisanes' content was accurately weighed on an Adam® PW254 analytical balance in a crucible taken previously to constant weight. It was incised gently at the beginning and then gradually increasing the temperature to 675 °C. The test was done for two hours. After this period, the ash was cooled in a desiccator and weighed in the same balance to obtain the total ash percentage. Later, it was placed in the Thermo Scientific® HeraTherm oven stove and dried for 30 minutes at 105 °C, cooled in the desiccator, and weight. These periods of heating and cooling were continued until reaching a constant weight. The compliant sample was considered if the total ash was less than 6.0% of the initial weight.<sup>[5]</sup>

#### **Acid-insoluble ash test**

The test was performed according to the general chapter <561> of the USP 42.<sup>[19]</sup> 25 ml of HCl 3 N were added to each sample obtained from the total ash test. After this, they were heated to boiling for 5 minutes. The insoluble material was transferred to a Boeco® grade 389 filter paper. It was then washed with hot water and transferred to a melting pot (brought previously to constant weight). Initially, it was incised gently and then gradually increased the temperature to 675 °C. The assay was made for two hours. After cooling in a desiccator, it was weighed again in the same balance. Later, it was placed in the Thermo Scientific® HeraTherm oven stove and dried for 30 minutes at 105 °C, cooled in the desiccator, and weight. These periods of heating and cooling were continued until reaching a constant weight. The acceptance specification is that the insoluble ash percentage should not be more than 4.0%.<sup>[5]</sup>

#### **Microbial enumeration tests**

The tests were executed according to the general chapter <61> of the USP 42.<sup>[19]</sup> 10 g of the product was taken and added in 100 ml of Bacto™ casein-soybean digest broth. Two Petri dishes were prepared for each medium at the required dilution level. The plates with Bacto™ casein-soybean digest agar were used for the total aerobic microbial count

(incubated at 33 °C for 48 hours). For yeasts and molds count, Liofilchem<sup>TM</sup> potato dextrose agar was employed (incubated at 22.5 °C for five days). Finally, the arithmetic mean of each medium's enumeration was taken, and the number of colony-forming units (CFU) calculated per g of product. According to the RTCA 11.03.56.09, the total aerobic microbial count should not exceed 10<sup>7</sup> CFU/g, and total combined yeasts and molds count should not exceed 10<sup>5</sup> CFU/g.<sup>[16]</sup>

### Microorganisms' specific tests

The procedures of chapter <2022> of USP 41 were followed.<sup>[19]</sup>

*Escherichia coli* absence test: 10 g of the product to be analyzed were added in 100 ml of Bacto<sup>TM</sup> casein-soybean digest broth. The sample was incubated at a temperature of 33 °C for 24 hours. Then, a 1.0 ml sample aliquot was tip out into a container with Difco<sup>TM</sup> MacConkey broth, mixed, and incubated at 44 °C for 48 hours. After that, two samples (1.00 ml of the broth) were taken to inoculate two BBL<sup>TM</sup> MacConkey agar plates. Plates were incubated at 33 °C for 24 hours. In the end, both plates were examined.

*Salmonella sp* absence test: from the sample preparation for the *E. coli* absence test, a 1.00 ml aliquot was taken and transferred to 10 ml of Difco<sup>TM</sup> Rappaport Vassiliadis Salmonella enrichment broth. The solution was mixed and incubated at 33 °C for 24 hours. After this, two samples were taken (1.00 ml) to inoculate two plates of Difco<sup>TM</sup> xylose lysine deoxycholate agar (incubated at 33 °C for 24 hours). Finally, both plates were examined.

Both tests complied if there was an absence of both pathogenic microorganisms.

## RESULTS AND DISCUSSION

**Table 1** includes the aspects stipulated for primary and secondary packaging, and the compliance of each batch of the commercial brands evaluated.

Regarding the primary packaging, none of the batches met the information required by the RTCA 11.04.41.06.<sup>[16]</sup>, precisely the batch number and the expiration date. The batch number is related to the product's traceability, referred to as the ability to follow a product's movement through all the stages involved in its production and distribution.<sup>[20]</sup> Although this number is included in the secondary packaging, the problem is that the pharmaceutical form (tisane) is generally consumed individually (they are kept in their primary packaging in other containers, not in its original box). In this way, it is not easy to make the respective report and take the appropriate actions with maximum efficiency and speed if a problem occurs.<sup>[20]</sup>

Additionally, the expiration date is the maximum date at which the product is still under its quality specifications. If a drug is consumed after this one, there is a possibility that its properties have changed.<sup>[21]</sup> Given this, consumers' ignorance of the expiration date puts them at risk since they could be using an unsafe and/or ineffective product.

**Table 1: Compliance with the labeling requirements of primary and secondary packaging by the evaluated batches of two cinnamon tea brands commercialized in Costa Rica.**

Requirement	Fulfillment					
	Brand 1			Brand 2		
	1	2	3	4	5	6
<i>Primary packaging</i>						
<b>Product name</b>	Yes	Yes	Yes	Yes	Yes	Yes
<b>Batch number</b>	No	No	No	No	No	No
<b>Expiration date</b>	No	No	No	No	No	No
<b>Manufacturer laboratory name or logo</b>	Yes	Yes	Yes	Yes	Yes	Yes
<i>Secondary packaging</i>						
<b>Product name</b>	Yes	Yes	Yes	Yes	Yes	Yes
<b>Pharmaceutical form</b>	Yes	Yes	Yes	Yes	Yes	Yes
<b>Indications</b>	Yes	Yes	Yes	Yes	Yes	Yes
<b>Employment form</b>	Yes	Yes	Yes	Yes	Yes	Yes
<b>Active ingredients, including quantity</b>	No	No	No	No	No	No
<b>Registration number</b>	Yes	Yes	Yes	Yes	Yes	Yes
<b>Manufacturer name and country of origin</b>	Yes	Yes	Yes	Yes	Yes	Yes
<b>Net amount of the finished product</b>	Yes	Yes	Yes	Yes	Yes	Yes
<b>Batch number</b>	Yes	Yes	Yes	Yes	Yes	Yes
<b>Storage conditions</b>	Yes	Yes	Yes	Yes	Yes	Yes
<b>Expiration date</b>	Yes	Yes	Yes	Yes	Yes	Yes
<b>Contraindications and warnings</b>	No	No	No	No	No	No
<b>Interactions</b>	No	No	No	No	No	No
<b>Side effects</b>	No	No	No	No	No	No
<b>General labeling</b>	No	No	No	No	No	No
<b>Specific labeling</b>	No	No	No	No	No	No
<b>Posology</b>	No	No	No	No	No	No
<b>Administration route</b>	No	No	No	No	No	No
<b>Use during pregnancy, breastfeeding, elderly, and children under 2 years</b>	No	No	No	No	No	No

Referring to the secondary packaging, it does not comply with the qualitative-quantitative composition. It is known that the active substances of the cinnamon bark are cinnamaldehyde (the most representative substance, which can be found up to 90%, depending on the extraction method) and eugenol.<sup>[22] [23]</sup> For this reason, their amount per tisane should be indicated, so that this specification is met. Two other requirements not shown in the package



related to the composition are the cinnamon's posology (dose allowed to achieve the beneficial effects; 1 to 3 g per day of product in 100 ml of water)<sup>[24]</sup> and the administration route. The information is necessary since the person needs the exact way to consume it to achieve the desired pharmacological effects without unnecessary risks.

The interactions are not mentioned either. Some studies have shown that cinnamon can interact with medications that low glucose levels,<sup>[25]</sup> including oral hypoglycemic agents.<sup>[7]</sup> Besides, due to coumarin's presence, caution should be exercised with people consuming antiplatelet or anticoagulant medications, and drugs capable of affecting the liver. Its concomitant use can increase the risk of bleeding and cause liver damage.<sup>[25]</sup> A study made in the murine liver *in vitro* suggests that cinnamon root components can interact with microsomal cytochrome 450, causing interactions with drugs metabolized by it.<sup>[26]</sup>

Furthermore, this product's warnings include that caution should be considered if consumed by people with diabetes and/or low blood pressure. Also, care should be taken because of some adverse effects, including diarrhea, vomiting, dizziness, and drowsiness. Likewise, the infusion oil can irritate the mucous membranes, including the stomach, intestinal, and urinary tracts.<sup>[27]</sup> In some cases, it has been linked to the appearance of states of nervousness.<sup>[28]</sup>

Other data not indicated in the secondary packaging were the general and the specific labeling. The general labeling includes phrases like "Keep out of reach of children" and "For over-the-counter modality: If symptoms persist, consult your doctor." Moreover, specific labeling is established for each product, and the information obtained should include this section.<sup>[16]</sup>

One last detail is the absence of information about its use during pregnancy, breastfeeding, elderly, and children under two years. This situation is worrisome. There is no evidence that large amounts of cinnamon intake could be safe for pregnant and breastfeeding women.<sup>[24]</sup>

Other tests performed were organoleptic tests, which refer to sensory evaluations.<sup>[29]</sup> In natural products, they assure medicinal plants' identity, purity, quality, and activity.<sup>[30]</sup> Various ethnobotanical studies have shown the relevance of these characteristics to distinguish between medicinal and non-medicinal plants. Even their selection and their indications can be explained by attributes such as taste and smell.<sup>[31]</sup> For the studied batches



of each brand of cinnamon tisanes, all met the specifications, supporting that it is cinnamon, despite being in the presence of a crush of the plant bark.

Another critical assessment is foreign matter. It is defined as any material from the medicinal plant or materials other than those specified.<sup>[32] [33]</sup> Therefore, within the control and quality standards, this matter must be removed.<sup>[34]</sup> For the foreign organ test carried out, leaves were found in the six batches (**Table 2**). This assay is required since the plant's medicinal effects vary according to its part.<sup>[35]</sup>

Additionally, in terms of foreign elements, plastic was found in two batches of brand 2, but the percentages obtained corresponded to 0. However, it is relevant to notice, because the raw material must be free of any non-plant component, including soil, insect parts, and animal excreta.<sup>[36]</sup> For the study made, all the batches followed the pharmacopoeial specifications.

**Table 2: Percentage of foreign organs and foreign elements for the three batches of each cinnamon tea brand commercialized in Costa Rica (batches 1 to 3 for brand 1 and 4 to 6 for brand 2).**

Batch	Total sample weight g	Foreign organs weight g	Foreign organs percentage %	Foreign elements weight g	Foreign elements percentage %
1	12.4477	0.0035	0	0	0
2	13.6670	0.0078	0	0	0
3	14.2953	0.0018	0	0	0
4	18.8990	0.0071	0	0.0006	0
5	17.9153	0.0034	0	0	0
6	18.1173	0.0089	0	0.0001	0

The minimum fill test was executed to know if the tisanes were appropriately filled. According to Costa Rican legislation, the test is accomplished only in the case of a complaint.<sup>[16]</sup> **Table 3** shows the minimum fill value obtained for each tisane, its labeling percentage, and the average for each batch of both brands.

**Table 3: Labeling percentage of each batch of two cinnamon tea brands commercialized in Costa Rica.**

Tisane	Weight g	Labeling percentage %	Weight g	Labeling percentage %	Weight g	Labeling percentage %
	Brand 1					
	Batch 1		Batch 2		Batch 3	
1	1.3865	107	1.3255	102	1.3734	106
2	1.3736	106	1.3871	107	1.3180	101
3	1.3606	105	1.3630	105	1.3645	105
4	1.4184	109	1.3472	104	1.3648	105
5	1.4337	110	1.3497	104	1.3897	107
6	1.4275	110	1.3743	106	1.3415	103
7	1.3396	103	1.3806	106	1.3790	106
8	1.3801	106	1.3822	106	1.3730	106
9	1.3726	106	1.3836	106	1.3350	103
10	1.3880	107	1.3666	105	1.3681	105
Mean	1.3881	107	1.3667	105	1.3607	105
	Brand 2					
	Batch 4		Batch 5		Batch 6	
1	1.8927	105	1.8413	102	1.8110	101
2	1.9186	107	1.7840	99	1.8051	100
3	1.8869	105	1.7753	99	1.8663	104
4	1.9094	106	1.8196	101	1.8254	101
5	1.9382	108	1.7655	98	1.8286	102
6	1.9020	106	1.8325	102	1.8705	104
7	1.8731	104	1.8431	102	1.7760	99
8	1.9519	108	1.8409	102	1.8847	105
9	1.8546	103	1.7664	98	1.8070	100
10	1.9330	107	1.8163	101	1.8912	105
Mean	1.9060	106	1.8085	100	1.8366	102

Regarding the minimum fill test, batches 1, 2, 3 of brand 1 were under the established pharmacopoeial standards,<sup>[19]</sup> obtaining average values of 107, 105, and 105%, respectively, concerning the labeling, which was 1.3 g. A similar situation happened with batches 4, 5, and 6 of brand 2, since values of 106, 100, and 102% were determined, respectively, about its labeling (1.8 g).

The test establishes a minimum that each tisane must contain, but not a maximum. This situation allows industries to add more amount without affecting their marketing permission. Still, in this investigation, more care was appreciated in the filling process regarding other researches made, where values were higher than 110%.<sup>[12] [13] [14] [15]</sup> The results show that producers are having greater control over the filling process (usually an acceptable

pharmacopoeial range for these products is 90 to 110%). In this way, it is ensured that the consumer receives a dose in the therapeutic range, which is defined as the concentration range at which substantial clinical benefits are obtained in the absence of unacceptable adverse effects.<sup>[37]</sup> In an overdose event, the person can endanger their health causing agitation, hypoglycemia, and tachycardia because of coumarin presence in the drink.<sup>[25]</sup>

Natural products also experience contamination, which can be accomplished by inappropriate human activities, such as heavy metals exposure. This scenario has severe consequences on people's health.<sup>[24]</sup>

One of the heavy metals is lead. Its exposure is mainly through the respiratory and gastrointestinal tracts. The amount absorbed by the intestine is around 10 to 15%, and this value increases if there are iron, zinc, or calcium deficiencies. Its toxic effects are occasioned by combining the sulfhydryl group in proteins (inhibiting sulfhydryl-dependent enzymes) and oxidative stress (increase in the oxidized form of glutathione or glutathione disulfide and inhibition of enzymes, such as superoxide dismutase and catalase).<sup>[38] [39]</sup>

As a consequence of this toxicity mechanism, the metal can generate failure in various body systems. People may have abdominal pain, anemia, hypertension, hearing impairment, peripheral neuropathy, and kidney failure.<sup>[40]</sup> Furthermore, it can produce proximal tubular damage, glomerular sclerosis, interstitial fibrosis, proteinuria, tubulointerstitial fibrosis, infertility, spontaneous abortion, stillbirth, premature birth, and low birth weight.<sup>[40] [41]</sup> Other consequences are intellectual, behavioral, or motor deficits, hand-eye coordination problems, reaction time difficulties, and poor performance on intelligence tests.<sup>[41]</sup>

Another compound is arsenic. Its poisoning occurs as a consequence of the consumption of improperly treated water.<sup>[42]</sup> Its toxicity mechanisms include the inhibition of cellular enzymes that contain sulfhydryl groups and the replacement of phosphate molecules in "high energy" compounds (arsenolysis). Trivalent arsenic compounds inhibit enzymes to a greater extent, while pentavalent ones are more involved in arsenolysis.<sup>[43]</sup>

Intoxication with arsenic compounds begins with nausea, vomiting, abdominal pain, and severe diarrhea.<sup>[42]</sup> Subsequently, prolonged exposure can originate tracheal and bronchogenic carcinomas, hepatic angiosarcomas, and various skin cancers. Myelogenous

leukemia, pigmentation changes, palmar and plantar hyperkeratosis, and anemia can also be developed.<sup>[43]</sup>

For these reasons, limit tests were done. The distinct batches evaluated of cinnamon's commercial brands showed to be below the permitted limits, fulfilling the quality requirements.

In addition, the quality of natural products is evaluated with the loss on drying test. Drying is an essential process of the raw material from which natural products are made, allowing their storage for a more extended period during their production and is considered a way to preserve their bioactivity and increase their useful life.<sup>[44]</sup> Another reason is the inhibition of microbial growth (closely related to the microbial enumeration tests discussed later) by minimizing growth (decreased water activity) and, therefore, contamination by bacteria, fungi, and yeasts.<sup>[45] [46]</sup> This technique prevents the development of specific biochemical reactions from microorganisms' metabolism, altering the product's organoleptic properties,<sup>[47]</sup> specifically its pharmacological utility. Likewise, raw materials become vulnerable to the presence of microbial toxins, including the aflatoxins, of great danger to humans (carcinogenic activity).<sup>[48]</sup>

For the two brands of tea, information regarding the loss of drying is observed in **Table 4**. From the specifications given,<sup>[19]</sup> all the batches are in accordance, obtaining a value of less than 12% for each one. The above information shows the producers' good work related to product elaboration.

Similar to the previous test, the assays for total ash and acid-insoluble ash were accomplished. Both allow to determine the presence of any foreign material, the inorganic composition of the raw material, and its purity,<sup>[49]</sup> mostly when they are crushed,<sup>[50]</sup> as happens with cinnamon teas. The total ash value includes metals, carbonates, phosphates, silicates, and silica found (including physiological and non-physiological ash). A high number indicates contamination, substitution, adulteration, or neglect during raw material preparation. Additionally, acid-insoluble ash represents contamination with silica (earth and sand, for example).<sup>[51] [52]</sup> The samples studied in this case showed the values indicated in **Table 5** for these tests.

**Table 4: Percentage of loss of drying for the three batches evaluated of each cinnamon tea brand commercialized in Costa Rica (batches 1 to 3 for brand 1 and batches 4 to 6 for brand 2).**

Batch	Initial weight g	Final weight g	Loss on drying percentage %
1	1.4056	1.2662	9.9
2	1.4025	1.2604	10.1
3	1.3954	1.2541	10.1
4	1.8132	1.6198	10.7
5	1.8033	1.6140	10.5
6	1.7864	1.6003	10.4

**Table 5: Percentage of total ash and acid-insoluble ash for the three batches evaluated of each cinnamon tea brand commercialized in Costa Rica (batches 1 to 3 for brand 1 and batches 4 to 6 for brand 2).**

Batch	Initial weight g	Total ash weight g	Total ash percentage %	Acid-insoluble ash weight g	Acid-insoluble ash percentage %
1	2.6842	0.0867	3.2	0.0027	0.1
2	2.7693	0.0845	3.1	0.0034	0.1
3	2.6177	0.0807	3.1	0.0094	0.4
4	3.7635	0.1415	3.8	0.0087	0.2
5	3.7063	0.1412	3.8	0.0052	0.1
6	3.5975	0.1209	3.4	0.0036	0.1

These tests indicate that all the batches of the two brands comply with total ash (the highest value obtained was 3.8%) and acid-insoluble ash specifications (the highest value was 0.4%). The results are evidence of the purity of the manufacturers' raw material utilized and the quality provided to consumers.

Other assays to determine the quality of natural products are related to the presence or absence of microorganisms. The growth of pathogenic microorganisms can affect the physicochemical characteristics of the product and must be controlled.<sup>[53]</sup> For the microbial enumerations of mesophilic microorganisms, fungi, and yeasts, the batches evaluated for both commercial cinnamon tea brands complied with the specifications established by the RTCA.<sup>[16]</sup> These results show the proper handling of the crude drug, because of the possibility of microbial contamination of water, soil, or animals during production and human activities (harvesting, drying, and classification).<sup>[54]</sup> The control of humidity through the drying process (shown above) is reflected in a decrease in microbial growth. If there is high

humidity in the packaged product, the water activity can increase to a level where microorganisms' growth damages the product. Besides, it can put the person in unnecessary danger.<sup>[55]</sup> Clinical cases showed significant infections assignable to the consumption of contaminated products.<sup>[56]</sup>

The last two analyses were the presence or absence of pathogenic Enterobacteriaceae, specifically *E. coli* and *Salmonella* sp. These microorganisms are of great importance due to their relationship with various health problems that affect mainly high-risk groups such as infants, pregnant women, immunosuppressed people, and the elderly.<sup>[57]</sup> *E. coli* is a commensal bacterium usually found as part of the intestinal microbiota of homothermic animals. Still, this species' pathogenic strains are capable of causing intestinal and extraintestinal diseases in humans.<sup>[58]</sup> For its part, *Salmonella* sp also belongs to the family of Enterobacteriaceae. More than 2,600 serotypes belonging to *Salmonella enterica* have been described, and many are capable of affecting human health.<sup>[59]</sup>

For this investigation, the absence of both pathogens was obtained in the different batches of cinnamon tea brands. This finding demonstrates that the population can safely consume them, especially since enteric diseases are leading causes of illness and death in developing countries.<sup>[60]</sup> It should be noted that the test for the absence of *Salmonella* sp. is not performed according to the Costa Rican regulations, as it does not apply to products that must be consumed with hot water. However, as indicated in previous works, in this country, people use hot water,<sup>[13] [14] [15]</sup> which is not capable of killing *Salmonella* sp.<sup>[61]</sup>

Finally, the reproducibility in terms of quality parameters was accomplished for the various tests implemented on the batches of each commercial brand. This finding is relevant because many parameters affect the requirements of the raw material, among them, the plant part employed, the harvest site (including altitude), the harvest periods (season and time of day), the drying method, and the exposure to pathogens, among others.<sup>[62] [63]</sup> There is significant care to provide products that maintain their quality, safety, and efficacy, regardless of the batch utilized.

## CONCLUSIONS

The three batches of each cinnamon tea brand commercialized in Costa Rica complied with the specifications established in the official documents for the following tests: organoleptic, foreign matter, minimum fill, heavy metals limit (lead), arsenic limit, loss on drying, total

ash, acid-insoluble ash, microbial enumerations, and specific microorganisms (*E. coli* and *Salmonella* sp.). The only one with nonconformities was the labeling test. For the primary packaging, the batch number nor the expiration date were found. As a complement, for the secondary one, the information regarding active ingredients (including quantity), contraindications and warnings, interactions, side effects, general and specific labeling, posology, administration route, and use during pregnancy, breastfeeding, elderly, and children under two years were absent.

Furthermore, there was reproducibility in terms of quality specifications for the batches of the same product. The results reflected the work accomplished by the manufacturers of these herbal teas to obtain quality products for Costa Rican consumers.

### ACKNOWLEDGMENTS

To the Laboratory of Pharmacognosy, the Laboratory of Phytopharmacology and Pharmaceutical and Cosmetic Technology (LAFITEC, for its Spanish acronym), the Laboratory of Pharmaceutical Products Analysis, and the Laboratory of Pharmaceutical Analysis and Assessment (LAYAFA, for its Spanish acronym) of the Universidad de Costa Rica, for allowing the use of different equipments and materials in this research.

### REFERENCES

1. Taylor WF, Jabbarzadeh E. The use of natural products to target cancer stem cells. *Am J Cancer Res*, 2017; 7(7): 1588-1605.
2. Yuan H, Ma Q, Ye L, Piao G. The Traditional Medicine and Modern Medicine from Natural Products. *Molecules*, 2016; 2(5): 559.
3. McKay DL, Blumberg JB. A Review of the Bioactivity of South African Herbal Teas: Rooibos (*Aspalathus linearis*) and Honeybush (*Cyclopia intermedia*). *Phytother Res*, 2007; 21(1): 1-16.
4. WHO Expert Committee on Specifications for Pharmaceutical Preparations. WHO Technical Report Series. Geneva; World Health Organization: 2018. Report No. 52.
5. World Health Organization. WHO monographs on selected medicinal plants – Volume 1. Geneva; World Health Organization: 1999.
6. Orwa C, Mutua A, Kindt R, Jamnadass R, Simons A. *Cinnamomum verum* [Internet]. Nairobi: World Agroforestry; 2009 [cited 2020 14 Apr]. Available from: [http://apps.worldagroforestry.org/treedb2/AFTPDFS/Cinnamomum\\_verum.PDF](http://apps.worldagroforestry.org/treedb2/AFTPDFS/Cinnamomum_verum.PDF)



7. Ranasinghe P, Pigera S, Sirimal Premakumara GA, Galappaththy P, Constantine GR, Katulanda P. Medicinal properties of 'true' cinnamon (*Cinnamomum zeylanicum*): a systematic review. *BMC Complement Altern Med*, 2013; 13: 275.
8. Ekor M. The growing use of herbal medicines: issues relating to adverse reactions and challenges in monitoring safety. *Front Pharm*, 2014; 4: 177.
9. Maldonado JC. Calidad, seguridad y eficacia de los medicamentos. In: López Salcedo D (ed.). *Acceso Público a Medicamentos de Calidad: Las Compras Públicas como Mecanismo para Garantizar el Derecho a la Salud*, Quito; Servicio Nacional de Contratación Pública: 2015, pp. 38-41.
10. Indrayanto G. Recent Development of Quality Control Methods for Herbal Derived Drug Preparations. *Nat Prod Commu*, 2018; 13(12): 1599-1606.
11. Jan S, Abbas N. *Himalayan Phytochemicals*. Cambridge; Elsevier: 2018.
12. Mora Román JJ, Alvarado Fernández MJ, Apú Leitón N, Arroyo Solórzano JD, Espeleta González D, Piedra Navarro H, et al. Pruebas Fisicoquímicas para la Evaluación de la Calidad de una Marca Costarricense de Tisanas de Valeriana. *Revista Médica de la Universidad de Costa Rica*, 2018; 12(1): 15-26.
13. Mora Román JJ, Bermúdez Carvajal K, Cordero Menéndez Y, Hernández Vega R, Porras León D, Vargas González R, et al. Importance of the Quality Control of Herbal Teas: Evaluation of two Senna Leafs Tisanes Brands Commercialized in Costa Rica through Physicochemical and Microbiological Assays Stipulated in the Central American Technical Regulation 11.03.56.09. *Sch Acad J Pharm*, 2018; 7(8): 372-380.
14. Mora Román JJ, Agüero Brenes N, Angulo Morales C, Castro Solís J, Hidalgo Carrillo G, van Hoof Gómez M, et al. Physicochemical and Microbiological Assays for Quality Evaluation of a Brand of *Mentha Piperita* Tisanes in Costa Rica Market: Employment of the Central American Technical Regulation. *Journal of Drug Delivery and Therapeutics*, 2018; 8(5): 329-337.
15. Gamboa Camacho S, González Vargas O, Guevara Saborío G, Murillo Castillo B, Loría Gutiérrez A, Blanco Barrantes J, et al. Quality Control of a Boldo Tisanes Brand Commercialized in Costa Rica Following the Central American Technical Regulation for Natural Products. *Saudi J Med Pharm Sci*, 2020; 6(1): 123-132.
16. Consejo de Ministros de Integración Económica Centroamericana. RTCA 11.03.56:09 (Reglamento Técnico Centroamericano de Productos Farmacéuticos. Productos Naturales Medicinales para Uso Humano. Verificación de la Calidad). San José; Consejo de Ministros de Integración Económica Centroamericana: 2011.

17. Consejo de Ministros de Integración Económica Centroamericana. RTCA 11.04.41:06 (Reglamento Técnico Centroamericano. Productos Farmacéuticos. Productos Naturales Medicinales para Uso Humano. Requisitos de Etiquetado) San José; Consejo de Ministros de Integración Económica Centroamericana: 2011.
18. European Pharmacopoeia Commission. European Pharmacopoeia 10th ed. Strasbourg; Council of Europe: 2019.
19. United States Pharmacopeia Convention. Pharmacopeia 42 and National Formulary 37. Maryland; United States Pharmacopeia Convention: 2019.
20. Agencia Española de Seguridad Alimentaria. Guía para la aplicación del sistema de trazabilidad en la empresa agroalimentaria. Madrid; Ministerio de Sanidad y Política Social: 2009.
21. Debesa García F, Fernández Argüelles R, Pérez Peña J. La caducidad de los medicamentos: justificación de una duda. *Rev Cubana Farm*, 2004; 38(3).
22. Nabavi SF, Di Lorenzo A, Izadi M, Sobarzo-Sánchez E, Daglia M, Nabavi SM. Antibacterial Effects of Cinnamon: From Farm to Food, Cosmetic and Pharmaceutical Industries. *Nutrients*, 2015; 7(9): 7729-7748.
23. Rao PV, Gan SH. Cinnamon: A Multifaceted Medicinal Plant. *Evid Based Complement Alternat Med*, 2014; 2014.
24. Drugs.com. Cinnamon [Internet]. Auckland: Wolters Kluwer Health. 2019 Jan 17 [cited 2020 Mar 21]. Available from: [https://www.rxlist.com/cinnamon\\_bark/supplements.htm](https://www.rxlist.com/cinnamon_bark/supplements.htm)
25. Martini N. Cinnamon. *J Prim Health Care*, 2015; 7(1): 77.
26. Çolak Ç, Elibol E, Demir T, Kivçak B. Overdose of Cinnamon Barks is the Cause of Poisoning in the Geriatric Patient: Case Report. *Journal of US-China Medical Science*, 2018; 15: 26-28.
27. RxList. Cinnamon Bark [Internet]. California: RxList. 2019 Sep 9 [cited 2020 Mar 21]. Available from: [https://www.rxlist.com/cinnamon\\_bark/supplements.htm](https://www.rxlist.com/cinnamon_bark/supplements.htm)
28. Barrionuevo E, Moreno D. Recetas líquidas saludables: Cremas, sopas, batidos, aguas e infusiones. Amat Editorial; Barcelona: 2016.
29. Patil SG, Wagh AS, Pawara RC, Ambore SM. Standard Tools for Evaluation of Herbal Drugs: An Overview. *The Pharma Innovation Journal*, 2013; 2(9): 60-65.
30. Upton R, David B, Gafner S, Glasl S. Botanical ingredient identification and quality assessment: strengths and limitations of analytical techniques. *Phytochem Rev*, 2019.

31. de Medeiros PM, Santos Pinto BL, do Nascimento VT. Can organoleptic properties explain the differential use of medicinal plants? Evidence from Northeastern Brazil. *J Ethnopharmacol*, 2015; 159: 43-48.
32. Silva PSC, Francisconi LS, Gonçalves RDMR. Evaluation of Major and Trace Elements in Medicinal Plants. *J Braz Chem Soc*, 2016; 27(12): 2273-2289.
33. European Directorate for the Quality of Medicines & HealthCare. Guide for the elaboration of monographs on herbal drugs and herbal drug preparations. Strasbourg; European Directorate for the Quality of Medicines & Health Care: 2007.
34. Agro and Food Processing. Setting up a Medicinal Herbs Extraction Unit. Gujarat; Government of Gujarat: 2017.
35. Mahmoud T, Gairola S. Traditional knowledge and use of medicinal plants in the Eastern Desert of Egypt: a case study from Wadi El-Gemal National Park. *J Med Plant Stud*, 2013; 1(6): 10-17.
36. Bele AA, Khale A. Standardization of Herbal Drugs: An Overview. *Int Res J Pharm*, 2011; 2(12): 56-60.
37. Perucca E, Pisani F. Dose-Response Relationship and Therapeutic Drug Monitoring. In: Lasagna L, Erill S, Naranjo SA (eds.). Dose response relationships in clinical pharmacology: proceedings of the Esteve Foundation Symposium III. Dose-response relationships in man. ICS808, Michigan; Elsevier Science Publishers B.V.: 1989, pp. 201-214.
38. Muñoz M. Determinación de plomo y cadmio en hierbas medicinales [thesis]. Buenos Aires; Universidad de Belgrano: 2009.
39. Shukla V, Shukla P, Tiwari A. Lead poisoning. *Indian J Med Spec*, 2018; 9(3): 146-149.
40. Flora G, Gupta D, Tiwari A. Toxicity of lead: A review with recent updates. *Interdiscip Toxicol*, 2012; 5(2): 47-58.
41. Donzelli G, Caducci A, Llopis-Gonzalez A, Verani M, Llopis-Morales A, Cioni L, et al. The Association between Lead and Attention-Deficit/Hyperactivity Disorder: A Systematic Review. *Int J Environ Res Public Health*, 2019; 16(3): 382.
42. Nurchi VM, Djordevic AB, Crisponi G, Alexander J, Bjørklund G, Aaseth J. Arsenic Toxicity: Molecular Targets and Therapeutic Agents. *Biomolecules*, 2020; 10(2): 235.
43. Hall AH. Chronic arsenic poisoning. *Toxicol Lett*, 2002; 128(1): 69-72.
44. Mediani A, Abas F, Tan CP, Khatib A. Effects of Different Drying Methods and Storage Time on Free Radical Scavenging Activity and Total Phenolic Content of *Cosmos caudatus*. *Antioxidants*, 2014; 3(2): 358-370.

45. Manzano Santana P, Quijano-Avilés M, Chóe-Guaranda I, Barragán Lucas A, Viteri Espinoza R, Martínez D, et al. Effect of drying methods on physical and chemical properties of *Ilex guayusa* leaves. Rev Fac Nac Agron Medellín, 2018; 71(3): 8617-8622.
46. Guiné RPF. The Drying of Foods and Its Effect on the Physical-Chemical, Sensorial and Nutritional Properties. Int J Food Eng, 2018; 4(2): 93-100.
47. Roshanak S, Rahimmalek M, Goli SAH. Evaluation of seven different drying treatments in respect to total flavonoid, phenolic, vitamin C content, chlorophyll antioxidant activity and color of green tea (*Camellia sinensis* or *C. assamica*) leaves. J Food Sci Technol, 2016; 53(1): 721-729.
48. Castillo L, Baltodano E, Ramírez N, Vargas R, Hanley G. Design of Experiments Assessment for the Determination of Moisture Content in Five Herbal Raw Materials in Tea Products. Borneo Journal of Pharmacy, 2020; 3(1): 22-35.
49. Chaudhari RK, Girase NO. Determination of soluble extractives and physico-chemical studies of bark of *Sesbania sesban* (L) Merr. J Chem Pharm Res, 2015; 7(8): 657-660.
50. Pande M, Mohammed KB. Comparative Physio-Chemical Evaluation of Different Brands of Lauha Bhasmas. Herb Med, 2019; 5(1): 3.
51. Magbool FF, Elnima EI, Shayoub ME, Hamedelniei EI, Alhassan MS. Pharmacognostic, Physicochemical Standardization and Phytochemical Analysis of *Quercus infectoria* galls. Am J Res Commun, 2018; 6(10): 1-17.
52. Alam F, us Saqib QJ. Pharmacognostic standardization and preliminary phytochemical studies of *Gaultheria trichophylla*. Pharm Biol, 2015; 53(12): 1711-1718.
53. Jin TZ. Current State of the Art and Recent Innovations for Antimicrobial Food Packaging. In: Juneja VK, Dwivedi HP, Sofos JN (eds.). Food Microbiology and Food Safety: Research and Development, New York; Springer Nature: 2017, pp. 349-372.
54. Nakajima K, Nonaka K, Yamamoto K, Yamaguchi N, Tani K, Nasu M. Rapid monitoring of microbial contamination on herbal medicines by fluorescent staining method. Lett Appl Microbiol, 2005; 40(2): 128-132.
55. Goverde M. Microbial Requirements and Testing of Primary Packaging. Pharmaceutical Microbiological Quality Assurance and Control: Practical Guide for Non-Sterile Manufacturing. In: Roesti D, Goverde M (eds.). New Jersey; John Wiley & Sons, Inc.: 2020, pp. 153-188.
56. Mukherjee PK. Quality Control Evaluation of Herbal Drugs: Evaluating Natural Products and Traditional Medicine. Cambridge; Elsevier Inc.: 2019.

57. Iwu CD, Okoh AI. Preharvest Transmission Routes of Fresh Produce Associated Bacterial Pathogens with Outbreak Potentials: A Review. *Int J Environ Res Public Health*, 2019; 16(22): 4407.
58. Beleza AJF, Maciel WC, Carreira AS, Bezerra WGA, Carmo CC, Havt A, et al. Detection of Enterobacteriaceae, antimicrobial susceptibility, and virulence genes of *Escherichia coli* in canaries (*Serinus canaria*) in northeastern Brazil. *Pesq Vet Bras*, 2019; 39(3): 201-208.
59. Jajere SM. A review of *Salmonella enterica* with particular focus on the pathogenicity and virulence factors, host specificity and antimicrobial resistance including multidrug resistance. *Vet World*, 2019; 12(4): 504-521.
60. Bublitz DC, Wright PC, Bodager JR, Rasambainarivo FT, Bliska JB, Gillespie TR. Epidemiology of Pathogenic Enterobacteria in Humans, Livestock, and Peridomestic Rodents in Rural Madagascar. *PLOS One*, 2014; 9(7): e101456.
61. World Health Organization. Boil Water. Geneva; World Health Organization: 2015.
62. Nagore DH. Development and Quality Control of Natural Products. Chhattisgarh; Evicenpub Publishing: 2018.
63. Sorkin BC, Kuszak AJ, Williamson JS, Hopp DC, Betz JM. The Challenge of Reproducibility and Accuracy in Nutrition Research: Resources and Pitfalls. *Adv Nutr*, 2016; 7(2): 383-389.