

THE PHARMACOLOGY, TOXICITY, AND PHYTOCHEMISTRY OF IPOMOEA CARNEA: AN OVERVIEW

Anjali Kumari Maurya*¹ and Prashant Kumar Singh²

¹Research Scholar, Saraswati Higher Education and Technical (SHEAT) College of
Pharmacy, Varanasi, U.P.

²Assistant Professor, Saraswati Higher Education and Technical (SHEAT) College of
Pharmacy, Varanasi, U.P.

Article Received on
03 May 2023,

Revised on 23 May 2023,
Accepted on 13 June 2023

DOI: 10.20959/wjpr202310-28668

*Corresponding Author

Anjali Kumari Maurya

Research Scholar, Saraswati
Higher Education and
Technical (SHEAT) College
of Pharmacy, Varanasi, U.P.

ABSTRACT

Ipomoea carnea is a plant commonly known as Besharam or Behaya tree. It belongs to the Convolvulaceae family and is native to America. The plant grows quickly and has spread widely in India. If pregnant animals eat this plant, it can lead to a lack of bonding between the mother and her offspring. In this article, we focus on controlling the uncontrolled growth of *Ipomoea carnea* and using it as a source of biogas along with cow dung cake. Different species of *Ipomoea* can be found in various parts of India. We have gathered information and recent advancements regarding the medicinal importance of *I. carnea*. The plant's extracts have shown antibacterial, antifungal, antioxidant, anticancer, anticonvulsant, immunomodulatory, antidiabetic, hepatoprotective, anti-inflammatory, anxiolytic, sedative, and wound healing properties. However, there have also been reports of some toxic effects. This study discusses the major phytochemicals associated with the plant's bioactivity. Researchers in phytotherapy might find this review article helpful, as *I. carnea* has the potential to be a valuable source for drug development.

KEYWORDS: *Ipomoea carnea*, phytoconstituent, Besharam, Behaya, Shameless, flavonoid, quercetin.

INTRODUCTION

Plants have been our natural companions since ancient times, providing us with food, clothing, shelter, and medicine. They are self-sufficient organisms that produce various chemicals called phytochemicals. These phytochemicals are not essential for the basic

functions of plants but serve additional purposes such as defense against pests, microbes, viruses, or other plants, as well as attracting animals for pollination or seed dispersal. Some of these compounds can even mimic the molecules found in our own cells, like hormones or signaling molecules, and have effects on our bodies [Wink et al., 2003]. Others perform the similar function of human metabolites, probably because of similar molecular target. Such as brassinolides are plant steroid hormones, which regulate cell division and cell development in the plant, and are structurally similar to human growth-regulating steroids [Grove et al., 1979]. Tracing the history of medical knowledge claims that plants are the root of medicine. Archaeological, anthropological and historical evidences support the use of medicinal plants by thousands of years ago [Sumner et al., 2000]

Ipomoea carnea is also known as besharm or behaya because it has been claimed that eating its leaves causes pregnant goats to lose their relationship with their mothers. In Western India, it is widely available. It flourishes along roadsides, riversides, and wetland edges. *Ipomoea carnea* is found in India in 60 different species on average. It is a member of the family Convolvulaceae. Their height typically ranges from 1 to 5 metres. In the manufacturing sector (paper manufacture), it is helpful. It is widespread throughout Rajasthan, Madhya Pradesh, Kota, Chandigarh, and Maharashtra. [Sharma et al., 2013]

***Ipomoea carnea*: basic features and traditional usage**

Scientific Classification

Kingdom- Plantae

Sub kingdom- Tracheobionta

Division- Spermatophyta

Subdivision- Magnoliophyta

Class- Magnoliopsida – Dicotyledons

Subclass- Asteridae

Order- solanales

Family- Convolvulaceae

Genus- *Ipomoea* Species- *carnea*

Local Name

Hindi- Beshram, Behaya,

English- Bush Morning glory

Oriya- Behayo

Marathi- Beshram

Bengali- Beshram

Phytochemical properties of *Ipomoea carnea*

The fundamental element responsible for a plant material's pharmacological and therapeutic activities is its phytochemical makeup. The existence of bioactive phytochemicals was suggested by a long history of traditional medical use, but most of these substances were unknown. L-rhamnose, D-fucose, D-chinovose (6-deoxy-D-glucose), D-glucose, convolvulinolate (11-hydroxy-pentadecane acid), jalapinolate (11-hydroxypalmitic acid), 7-hydroxy-decane acid, and ipurolic acid (3, 11-dihydroxy-tetradecane acid) were found in the latex of *I.* In their report on the phytochemical analyses of *I. carnea* leaves, Tirkey et al. [Tirkey et al., 1988] discovered that the dried powdered leaves of the plant contain alkaloids, reducing sugars, glycosides, and tannins, while the latex of the plant contains acacetin-7-galactoside, flavone glycoside, and an unidentified saponin known as ipomotocin.

According to recent studies, *I. carnea*'s leaves, stem, and flowers contain a respectable quantity of polyphenols (30-70 mg of catechol equivalent per g of dry material) and flavonoids (80-120 mg of quercetin equivalent per g of dry material) [Khawar et al., 2010]. Alkaloids, carbohydrates, tannins, phenolic chemicals, proteins and amino acids, terpenoids and sterols, and saponins were all found in the methanol extract of *I. carnea*'s leaves and flowers, according (Arora et al., 2013). Only the terpenoid in the leaves, flowers, and seeds, as well as alkaloids of the polyhydroxylated class, have been partially characterised among these phytochemicals.

Chromatographic study on the leaves, flowers and seeds resulted in the isolation of swainsonine, 2-epi-lentiginosine, calystegines B1, calystegines B2, calystegines B3, calystegines C1 and N-methyl-trans-4-hydroxy-L-proline at varying combination and concentration [Adsul et al., 2009].

In case of protein, only one protein molecule has been isolated and characterized from the latex of *I. carnea*, known as "Carnein", which is a serine protease with a molecular weight of 80.24 kDa [Patel et al., 2007].

Ipomoea carnea contain variety of bioactive components such as phenolic acid, alkaloids, flavonoids, coumarins and sterols1-4

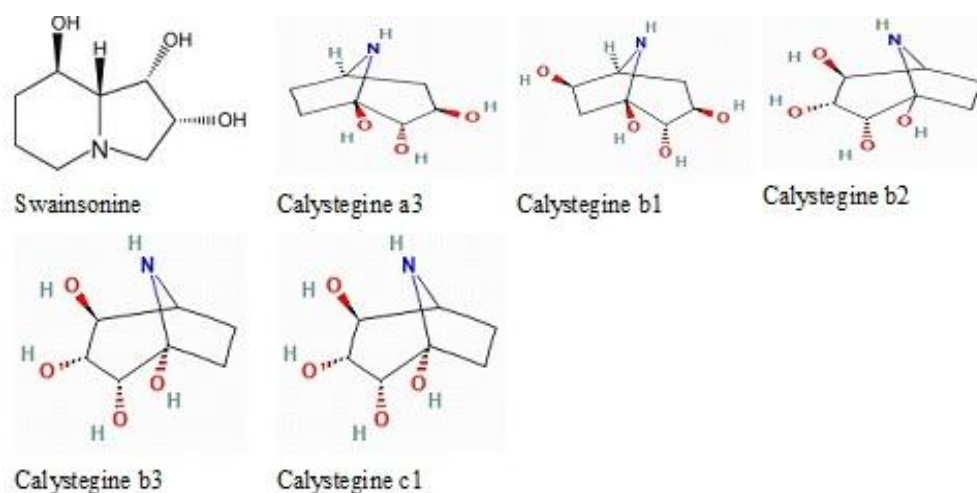


Figure 1: Major bioactive constituents of *Ipomoea carnea*. (A) Swainsonine, (B) Calystegine a3, (C) Calystegine b1, (D) Calystegine b2, (E) Calystegine b3 and (F) Calystegine c1. Source: Pubchem: www.pubchem.ncbi.nlm.nih.gov

The National Institute of Standard and Technology (NIST) database 2005 was used for the GC-MS analysis of the methanol extract of the leaf powder to determine the chemicals contained. By comparing the molecular weight and retention period, the spectra of unknown compounds were compared to those of recognised compounds kept in the NIST collection. Leaf powder included 22 bioactive phytochemical substances that were discovered. The majority of the substances are phenolic compounds, derivatives of flavonoids, carbohydrates, glycosides, saponins, and phytosterols based on factors such peak area percentage, molecular formula, and molecular weight. It has been discovered that these various active phytochemicals have a variety of properties that may aid in the prevention of various diseases. 2018 [Kar et al.]. A gas chromatography–mass spectrometry study on the hexane extract of *I. carnea* showed the presence of a panel of 13 compounds including hexadecanoic acid, stearic acid, 1,2-diethyl phthalate, n-octadecanol, octacosane, hexatriacontane, tetraacontane and 3-diethylamino-1-propanol [Haraguchi et al., 2003].

Medicinal/pharmacological action of *Ipomoea carnea* Over the last few decades, a number of researches demonstrated several researchers have been found to be antiinflammatry, antioxidant, antidiabetic, Anti-hepatotoxic, oxidative stress, liver protective etc [Khalid et al., 2011, Adsul et al., 2012, Abdul et al., 2012, Gupta et al., 2013, Khan et al., 2015, Ambiga et al., 2015, Gupta et al., 2012]. Other pharmacological activities are discussed below under.

REPORTED PHARMACOLOGICAL ACTIVITIES

Anti-microbial activity

I. carnea has been studied for its ability to fight bacteria and fungi. In one study [Adsul et al., 2012], it was found that the acetone extract of *I. carnea* is effective against *Proteus vulgaris* and *Salmonella typhimurium* bacteria, while the ethanol extract works against *Pseudomonas aeruginosa*. Another compound called dibutyl phthalate, found in *I. carnea*, has anti-bacterial properties and can fight certain types of gram negative bacteria like *Klebsiella pneumonia*, *Proteus mirabilis*, and *P. aeruginosa* [Khatiwora et al., 2012]. Some resin glycosides from *I. carnea* have been shown to enhance the effectiveness of antibiotics such as tetracycline, kanamycin, and chloramphenicol, which are commonly used to treat bacterial infections [Corona-Castañeda et al., 2012]. These resin glycosides may work together with other compounds in *I. carnea* to fight bacteria.

In traditional medicine, *I. carnea* has been used for treating skin diseases, but there is limited scientific research on this topic. However, a study by Mogle et al. [2013] demonstrated that the leaf extracts of *I. carnea* have antifungal properties and can inhibit the growth of fungi such as *Aspergillus niger*, *Penicillium digitatum*, *Botrytis cinera*, *Rhizopus arrhizus*, *Aspergillus flavus*, *Chaetomium brasiliense*, and *Rhizoctonia solani*. Among these fungi, the extract showed the strongest effect against *Aspergillus niger*. The main anti-fungal components in the leaves of *I. carnea* were found to be two coumarate isomers: (E)-octadecyl p-coumarate and (Z)-octadecyl p-coumarate [Nidiry et al., 2011].

Anti-oxidant activity

A class of compounds known as antioxidants can stop the oxidation of other molecules by squelching reactive free radicals. As a result, they may have positive health benefits on the prevention of degenerative diseases. According to [Khatiwora et al., 2010], the leaves, stem, and flowers of *I. carnea* contain significant amounts of antioxidants such as polyphenols and flavonoids. According to Adsul et al. (2012), *I. carnea* contains high levels of polyphenols and flavonoids that have been shown to have potent DPPH radical scavenging action. Particularly, this plant's blossom has higher concentrations of phytoconstituents that fight free radicals.

Anti-cancer activity

Hexane, chloroform, and ethyl acetate all had cytotoxic effects on the *I. carnea* fraction, with LC50 values of 141.4 g/mL, 211.28 g/mL, and 307.28 g/mL, respectively [Sharma et al.,

2013]. Isolated from *I. carnea*, the natural alkaloid swinsonine has been shown to have anti-cancer effects on a number of rat cancer models and human carcinoma. The apoptosis that is induced by swansonine in the human lung cancer cell line A549 prevents cell proliferation. The *I. carnea* alkaloid swainsonine was found to induce apoptosis in A549 cells by upregulating Bax, downregulating Bcl-2, promoting Bax translocation to mitochondria, triggering the mitochondria-mediated apoptotic pathway, releasing cytochrome C, and activating caspase-9 and caspase-3 [Li et al., 2012].

Anti-convulsant activity

The anti-convulsant activity of both polar and nonpolar extract was evaluated in mice and rats using the pentylenetetrazole and maximal electroshock (MES)-induced seizure models by Rout et al.^[30] The result of MES-induced convulsion showed that the polar extract significantly reduced extensor phase and stupor phase at a dose ranging from 200 mg/kg to 400 mg/kg. Indeed, the anti-convulsant activity was comparable to that of standard drug, Phenytoin. The polar extract also delayed the onset of time and increased the duration of pentylenetetrazole-induced convulsion. Thus extracts caused a significant dose-dependent increase in onset of convulsion compared with the control in pentylenetetrazole and MES-induced seizures [Rout et al., 2013].

Immunomodulatory effect

Only ten years ago, a study by Hueza et al. brought *I. carnea*'s immunomodulatory function into sharper focus [Hueza et al., 2003]. They claimed that a small dose of *I. carnea*'s aqueous fraction caused macrophages to phagocytose and produce hydrogen peroxide. Swainsonine, an alkaloid found in *I. carnea*, also exhibits immunomodulatory properties. Swainsonine typically functions by preventing glycoprotein metabolism. If rats were exposed to swinsonine when they were young, during breastfeeding, swinsonine modifies immunological function in adult rats [Hueza et al., 2011].

Anti-diabetic effect

The anti-hyperglycemic effect of aqueous extract of *I. carnea* leaves in streptozotocin-induced diabetes in rats was studied [Khalid et al., 2011]. A significant effect was observed by the extracts at 500 mg/kg dosage that was comparable to Glibenclamide (10 mg/kg). The phytoconstituents probably explains such anti-diabetic or hypoglycemic effect. The polyhydroxylated nortropane alkaloids calystegines are found in *I. carnea* and other Convolvulaceae plants.^[34] Calystegines B1 and C1 are potent competitive inhibitors of

bovine, human and rat β glucosidase activities. Calystegine B2 is a strong competitive inhibitor of the α -galactosidase activity in the livers of bovine, human and rat, while calystegines A3 and B2 are selective inhibitors of rat liver β -glucosidase [Asano et al., 1997].

Liver protective

Aqueous extract of *I. carnea* leaves was reported to restore the liver structure and functioning suggesting indicators (activity and/or expression) in a dose-dependent manner in a rat model of hepatotoxicity caused by carbon tetrachloride. The anti-oxidant properties of *I. carnea* are partially responsible for the hepatoprotective effect. The lipid peroxidation in liver tissue was said to be reduced by leaf aqueous extract, and the activity of anti-oxidant enzymes such as superoxide dismutase and catalase were said to be restored to normal levels. According to histological findings [Gupta et al., 2012], hepatocellular necrosis had also improved and the infiltration of inflammatory cells had decreased.

Anti-inflammatory effect

In formalin (0.1%)-induced rat paw edoema model, the anti-inflammatory efficacy of methanolic and petroleum ether extracts of *I. fistulosa*, a subspecies of *I. carnea*, leaves was reported. The extract appears to have anti-inflammatory effect at different acute phases of inflammation, according to a time-dependent evaluation of that activity. The presence of sitosterol in *I. fistulosa* or *I. carnea* may be responsible for this anti-inflammatory effect [Ruchi et al., 2009]. The ability of *I. carnea* leaf extracts to reduce inflammation was tested using a carrageenan (0.1%)-induced rat paw edoema model. This study's findings are comparable to those of the previous study, but more notably, the anti-inflammatory impact is developed in the early stages of inflammation [Khalid et al., 2011].

Anxiolytic activity

I. carnea seems to belong to the class of central depressants known as sedative-hypnotics. The elevated plus maze, open field test, and hole-board test model were used to examine the anxiolytic effect of the aqueous and methanolic extract of *I. carnea* leaves in mice [Bidkar et al., 2012]. The elevated plus maze is a typical behavioural paradigm because it may be used to research anxiety in animals in pharmacological, physiological, and behavioural approaches [Fabian et al., 2009]. Both the open field test and the hole-board test typically show emotional and/or anxious reactions to stress and indicate exploratory behaviour [Rodriguez et al., 1987; Takeda et al., 1998]. The potential of the aqueous and methanolic extract of *I. carnea* leaves to lessen anxiety is supported by each of these behavioural paradigms [Bidkar

et al., 2012]. Many nortropane alkaloids, notably calystegines B1, B2, C3, and the indolizidine alkaloid swainsonine, are present in *I. carnea*, and these alkaloids are assumed to have anxiolytic characteristics [de Balogh et al., 1999; Haraguchi et al., 2003].

Sedative activity

The sedative effect of the petroleum ether, alcohol and aqueous extracts of *I. carnea* leaf was evaluated in mice and rats using phenobarbitone-induced sleeping time and head dip test.^[30] Study reported that the duration of sleeping time in phenobarbitone-induced experimental models was increased in a dose-dependent manner with a significant decrease in locomotor activity at high dose, and in case of head dip test, exploratory behavioral potential was found to be decreased due to high dose of alcoholic and aqueous extract [Rout et al., 2013].

Wound healing activity

The flavonoids (Kaempferol and Kaempferol-3-O- β -D-glucoside) extracted from *I. carnea* flowers demonstrated considerable wound healing activity in the study using the incision and excision wound model [Ambiga et al., 2007]. Both the inflammatory and proliferative phases of wound healing are strongly influenced by these two flavonoids, according to macroscopic, biochemical, and histological aspects [Stadelmann et al., 1998]. Less macrophage, which are the predominant inflammatory cell type cells at the wound site, are one of the histological aspects of healing that signified the improvement of the inflammatory phase in another study [Rodero et al., 2010]. Interestingly, the improvement of the proliferative phase of wound healing was confirmed by a combination of macroscopic features and biochemical features, including an increase in granulation tissue and hydroxyproline concentration, respectively [Midwood et al., 2004]. Depending on the flavonoid kinds and wound types, different biochemical, histological, and macroscopic characteristics are present to different degrees.

Toxicity effects of *Ipomoea carnea*

General toxicity A number of studies reported the toxicological effects of *I. carnea*, mainly in goats and sheeps. Chronic ingestion of *I. carnea* has been reported to cause general weakness, loss of body weight, loss of hair, locomotor disturbance, loss of reflexes, intero-hepato-nephropathy, muscle tremors, ataxia, posterior paresis, paralysis and even death [Idris et al., 1973, Damir et al., 1987]. In Wister rats, some of the biochemical changes, like leukocytosis, anemia, an increase in serum aspartate Amino Transferase activity and decrease of albumin level have been noticed after *I. carnea* treatment [Amna et al., 2011]. The dihydroxynortropane alkaloids are thought to be responsible for these toxic effects of *I.*

carnea [Asano et al., 2001]. Also, it is suspected that the calystegines might act as coadjuvants of swainsonine in *I. carnea* toxicosis [Hueza et al., 2005].

Lysosomal storage disease induction

Lysosomal malfunction causes a category of uncommon inherited metabolic illnesses known as lysosomal storage disease [Winchester et al., 2000]. A small number of lysosomal storage diseases are caused by environmental causes, such as ingesting poisonous plants like locoweeds (*Astragalus* and *Oxytropis* spp.) [Van Kampen et al., 1969]. The majority of these diseases are hereditary abnormalities. According to a report by de Balogh et al. (1999), *I. carnea* causes lysosomal storage disease in goats. Ataxic, with head tremors and nystagmus, the affected animals were discovered. Lysosomal storage disorder is hypothesised to be caused by the glycosidase inhibitor phytoconstituents of *I. carnea*, notably swainsonine and calystegines.

Teratological property

Oral administration of the plant extract from days 6 to 20 of gestation was used to assess the effects of prenatal administration of *I. carnea* on pregnant rats and their progeny. According to the findings of that investigation, the thyroid, pancreas, liver, and kidneys of kids had organ-specific *I. carnea* toxicosis, which was characterised by cytoplasmic vacuolization. *I. carnea* treatment resulted in pups losing weight, having their thymus atrophy, and having their spleens grow larger [Hueza et al., 2003]. It has been demonstrated that feeding the offspring an aqueous extract of *I. carnea* did not result in severe developmental changes that resulted in behavioural changes [Schwarz et al., 2003]. In contrast, a different study by Schwarz et al. [Schwarz et al., 2007] demonstrated that giving *I. carnea* aqueous extract to animals causes a considerable reduction in 3,4-dihydroxyphenylacetic acid levels and an uptick in vanilmandelic acid levels in the striatum, cortex, and hypothalamus in a diffuse way. This demonstrated the pups' increased norepinephrine activity and decreased dopamine activity [Schwarz et al., 2007].

CONCLUSION

Phytomedicine has become popular again. It refers to using plants as medicine. Modern medicine is interested in traditional medicine, especially phytomedicine, alongside chemical medicine. In fact, many chemical medicines are derived from plants. Phytotherapy, which is using plant-based treatments, is believed to be an important approach in the future because it's safe and effective. *I. carnea* is a medicinal plant that has been used for thousands of years,

but there haven't been many scientific studies on it until recently. Scientists have started to show interest in it. However, most of the research on *I. carnea* has been done in labs and on animals. To develop it as a drug for humans, more studies need to be done, including pre-clinical and clinical trials. This review focuses on the basic research of *I. carnea* and its active components using animal models that mimic specific diseases like diabetes, immune deficiency, and cancer. Lab experiments using animal cells can help identify the specific effects of *I. carnea* on different organs and evaluate its safety. It's important to evaluate the effects of *I. carnea* on normal and diseased human cells to establish its potential as a treatment. Before *I. carnea* or its active components can be used clinically, they need to be evaluated for safety. It's hopeful that active compounds in *I. carnea* can be identified and used in clinical trials for drug development. More scientific studies, both clinical and basic, can establish *I. carnea* as an important plant in modern phytotherapy research.

REFERENCES

1. Abdul Latif K, Prasad AK, Kumar S, Iyer SV, Patel HA and Patel JA. Comparative antidiabetic studies of leaves of *Ipomoea carnea* and *Grewia asiatica* on streptozotocin induced diabetic rats. *Int. J. Pharm. Biol. Archives*, 2012; 3(4): 853-857.
2. Adsul V, Khatiwora E, Deshpande NR. Evaluation of antioxidant activity of *Ipomoea carnea* leaves. *J. Nat. Prod. Plant Resour*, 2012; 2(5): 584-588.
3. Adsul V, Khatiwora E, Kulkarni M, Tambe A, Pawar P, Deshpande N-. GC-MS study of fatty acids, esters, alcohols from the leaves of *Ipomoea carnea*. *Int J Pharm Tech Res.*, 2009; 1: 1224–6.
4. Adsul VB, Khatiwora E, Torane R, Deshpande NR. Anti-microbial activities of *Ipomoea carnea* leaves. *J Nat Prod Plant Resour*, 2012; 2: 597–600.
5. Alam S and Chowdhury SA. Pharmacological investigations of different extracts from the leaf of *Ipomoea fistulosa* (Family: Convolvulaceae). *Jordan J. Pharm. Sci.*, 2015; 8(3): 207-215.
6. Ambiga S and Jeyaraj M. Evaluation of in vitro Antioxidant Activity of *Ipomoea carnea* Jacq. *Int. J. Curr. Microbiol. App. Sci.*, 2015; 4(5): 327-338.
7. Ambiga S, Narayanan R, Gowri D, Sukumar D, Madhavan S. Evaluation of wound healing activity of flavonoids from *Ipomoea carnea* jacq. *Anc Sci Life.*, 2007; 26: 45–51.
8. Amna AA, Abdelgadir EH, Adam SE. Toxic effect of *Ipomoea carnea* leaves on wistar rats. *J Pharmacol Toxicol*, 2011; 6: 18–23.

9. Arora S, Kumar D, Shiba. Phytochemical, anti-microbial and anti-oxidant activities of methanol extract of leaves and flowers of *Ipomoea cairica*. *Int J Pharm Pharm Sci.*, 2013; 1: 198–202.
10. Asano N, Kato A, Matsui K, Watson AA, Nash RJ, Molyneux RJ, et al. The effects of calystegines isolated from edible fruits and vegetables on mammalian liver glycosidases. *Glycobiology*, 1997; 7: 1085–8.
11. Asano N, Yokoyama K, Sakurai M, Ikeda K, Kizu H, Kato A, et al. Dihydroxynortropane alkaloids from calystegine producing plants. *Phytochemistry*, 2001; 57: 721–6.
12. Bidkar JS, Bhujbal MD, Ghanwat DD, Dama GY. Anxiolytic activity of aqueous and methanolic extracts of *Ipomoea carnea* leaves. *Int J Univ Pharm Bio Sci.*, 2012; 1: 1–11.
13. Corona-Castañeda B, Pereda-Miranda R. Morning glory resin glycosides as modulators of anti-biotic activity in multidrugresistant gram-negative bacteria. *Planta Med.*, 2012; 78: 128–31.
14. Damir HA, Adam SE, Tartour G. Effects of *Ipomoea carnea* on goats and sheep. *Vet Hum Toxicol*, 1987; 29: 316–19.
15. De Balogh KK, Dimande AP, van der Lugt JJ, Molyneux RJ, Naudé TW, Welman WG. A lysosomal storage disease induced by *Ipomoea carnea* in goats in Mozambique. *J Vet Diagn Invest*, 1999; 11: 266–73.
16. Fabiana CV, Roseli S, Alexandre GP. Anxiolytic-like effect of *Sonchus oleraceus* L. in mice. *J Ethnopharmacol*, 2009; 124: 325–7.
17. Grove MD, Spencer GF, Rohwedder WK, Mandava N, Worley JF, Warthen JD, et al. Brassinolide, a plant growth-promoting steroid isolated from *Brassica napus* pollen. *Nature*, 1979; 281: 216–17.
18. Gupta RK, Chaudhary S, Vaishali, Singh Rk. Antihepatotoxic influence of aqueous extract of *Ipomoea carnea* against carbon tetrachloride induced acute liver toxicity in experimental rodents. *Asian J. Pharm. Cli. Res.*, 2012; 5(4): 262-265.
19. Gupta RK, Gupta AK, Swain SR, Vaishali, Gupta G, Khalid S, Suresh DK and Singh RK. Anti-hepatotoxic and antioxidant influence of *Ipomoea carnea* against anti-tubercular drug induced acute hepatopathy in experimental rodents. *J. Coast. Life Med.*, 2013; 1(4): 293-299.
20. Haraguchi M, Gorniak SL, Ikeda K, Minami Y, Kato A, Watson AA, et al. Alkaloidal components in the poisonous plants, *Ipomoea carnea* (Convolvulaceae). *J Agric Food Chem.*, 2003; 51: 4995–5000.

21. Haraguchi M, Gorniak SL, Ikeda K, Minami Y, Kato A, Watson AA, et al. Alkaloidal components in the poisonous plants, *Ipomoea carnea* (Convolvulaceae). *J Agric Food Chem.*, 2003; 51: 4995–5000.
22. Hueza IM, Dagli ML, Górnaiak SL, Paulino CA. Toxic effects of prenatal *Ipomoea carnea* administration to rats. *Vet Hum Toxicol*, 2003; 45: 298–302.
23. Hueza IM, Fonseca ES, Paulino CA, Haraguchi M, Górnaiak SL. Evaluation of immunomodulatory activity of *Ipomoea carnea* on peritoneal cells of rats. *J Ethnopharmacol*, 2003; 87: 181–6.
24. Hueza IM, Górnaiak SL. The immunomodulatory effects of *Ipomoea carnea* in rats vary depending on life stage. *Hum Exp Toxicol*, 2011; 30: 1690-700.
25. Hueza IM, Guerra JL, Haraguchi M, Naoki A, Górnaiak SL. The role of alkaloids in *Ipomoea carnea* toxicosis: a study in rats. *Exp Toxicol Pathol*, 2005; 57: 53–8.
26. Idris OF, Tartour G, Adam SE, Obeid HM. Toxicity to goats of *Ipomoea carnea*. *Trop Anim Health Prod.*, 1973; 5: 119–23.
27. Kar GBA, Rout SK, Mishra D. Phytochemical screening and GC-MS analysis of methanol extract of the leaves of *ipomoea carnea*. *World Journal of Pharmaceutical Research*, 2018; 7(13): 887-895.
28. Khalid MS, Singh RK, Kumar SJ, Suresh DK, Srinivas RK, Raddy NI. Anti-diabetic activity of aqueous extract of *Ipomoea carnea* leaves in streptozotocin induced diabetic rats. *Int J Pharmacol Bio Sci.*, 2011; 5: 45–54.
29. Khalid MS, Singh RK, Reddy IV, Kumar SJ, Kumar BS, Kumar GN, et al. Anti-inflammatory activity of aqueous extract of *Ipomoea carnea* jacq. *Pharmacology (Online)*, 2011; 1: 326–31.
30. Khalid MS, Singh RK, Reddy IVN, Kumar SJ, Kumar BS, Kumar GNS and Rao KS. Anti-inflammatory activity of aqueous extract of *Ipomoea Carnea* jacq. *Pharmacologyonline*, 2011; 1: 326-331.
31. Khan TY, Raina R, Verma PK, Sultana M, Mahrukh A. Protective effect of *Ipomoea carnea* Jacq leaves extracts on streptozotocin-induced oxidative stress in rats. *J. Exp. Integ. Med.*, 2015; 5(1): 1-5.
32. Khatiwora E, Adsul VB, Kulkarni M, Deshpande NR, Kashalkar RV. Anti-bacterial activity of dibutyl phthalate: a secondary metabolite isolated from *Ipomoea carnea* stem. *J Pharm Res.*, 2012; 5: 150–2.

33. Khatiwora E, Adsul VB, Kulkarni MM, Deshpande NR, Kashalkar RV. Spectroscopic determination of total phenol and flavonoid contents of *Ipomoea carnea*. *Int J ChemTech Res.*, 2010; 2: 1698–701.
34. Legler G. Die bestandteile des giftigen glykosidharzes aus *Ipomoea fistulosa* Mart. ex Choisy. *Phytochemistry*, 1965; 4: 29–41.
35. Li Z, Xu X, Huang Y, Ding L, Wang Z, Yu G, et al. Swainsonine activates mitochondria-mediated apoptotic pathway in human lung cancer A549 cells and retards the growth of lung cancer xenografts. *Int J Biol Sci.*, 2012; 8: 394–405.
36. Midwood KS, Williams LV, Schwarzbauer JE. Tissue repair and the dynamics of the extracellular matrix. *Int J Biochem Cell Biol.*, 2004; 36: 1031–7.
37. Mogle UP. Efficacy of leaf extracts against the post harvest fungal pathogens of cowpea. *Biosci Discov.*, 2013; 4: 39–42.
38. Nidiry ES, Ganeshan G, Lokesh A. Anti-fungal activity and isomerization of octadecyl p-coumarates from *Ipomoea carnea* subsp. *fistulosa*. *Nat Prod Commun*, 2011; 6: 1889–92.
39. Patel AK, Singh VK, Jagannadham MV. Carnein, a serine protease from noxious plant weed *Ipomoea carnea* (Morning Glory). *J Agric Food Chem.*, 2007; 55: 5809–18.
40. Rodero MP, Khosrotehrani K. Skin wound healing modulation by macrophages. *Int J Clin Exp Pathol*, 2010; 3: 643–53.
41. Rodriguez EL, Broitman ST, Foscolo MR. Effect of chronic ingestion of chlorimipramine and desipramine on the hole board response to acute stresses in male rats. *Pharmacol Biochem Behav*, 1987; 26: 207–10.
42. Rout SK, Kar DM. Sedative, anxiolytic and anti-convulsant effects of different extracts from the leaves of *Ipomoea carnea* in experimental animals. *Int J Drug Dev Res.*, 2013; 5: 232–43.
43. Ruchi J, Niles J, Surendar J. Evaluation of anti-inflammatory activity of *Ipomoea fistulosa* linn. *Asian J Pharm Clin Res.*, 2009; 2: 64–7.
44. Schwarz A, Górniak SL, Bernardi MM, Dagli ML, Spinoso HS. Effects of *Ipomoea carnea* aqueous fraction intake by dams during pregnancy on the physical and neurobehavioral development of rat offspring. *Neurotoxicol Teratol*, 2003; 25: 615–26.
45. Schwarz A, Hosomi RZ, Flório JC, Bernardi MM, Górniak SL, Spinoso HS. Rats offspring exposed to *Ipomoea carnea* and handling during gestation: neurochemical evaluation. *Braz Arch Biol Technol*, 2007; 50: 425–33.

46. Sharma A, Bachheti RK. A Review on *Ipomoea carnea*, International Journal of Pharm and Bio Sciences, 2013; 4(4): 363-377.
47. Sharma N, Gupta PC, Singh A, Rao CV. Brine shrimp bioassay of *Pentapetes phoenicea* linn. and *Ipomoea carnea* jacq. leaves. Der Pharmacia Lettre, 2013; 5: 162–7.
48. Stadelmann WK, Digenis AG, Tobin GR. Physiology and healing dynamics of chronic cutaneous wounds. Am J Surg, 1998; 176: 26S–38S.
49. Sumner J. The natural history of medicinal plants. Portland: Timber Press, 2000; 15–7.
50. Takeda H, Tsuji M, Matsumiya T. Changes in head dipping behavior in the hole board test reflects the anxiogenic and/ or anxiolytic state in mice. Eur J Pharmacol, 1998; 350: 21–9.
51. Tirkey K, Yadava RP, Mandal TK, Banerjee NL. The pharmacology of *Ipomoea carnea*. Indian Vet J., 1988; 65: 206–10.
52. Van Kampen KR, James LF. Pathology of locoweed poisoning in sheep. Pathol Vet, 1969; 6: 413–23.
53. Winchester B, Vellodi A, Young E. The molecular basis of lysosomal storage diseases and their treatment. Biochem Soc Trans, 2000; 28: 150–4.
54. Wink M. Evolution of secondary metabolites from an ecological and molecular phylogenetic perspective. Phytochemistry, 2003; 64: 3-19.