FARMACOGNOSTIC STUDY OF LEAF OF HYPTIS SUAVEOLENS L.

S. JELANI AND M. PRABHAKAR

Department of Botany, Osmaniya University, Hyderabad – 500 007, India.

Received: 29 May, 1991

Accepted: 15 June, 1991

ABSTRACT: This paper deals with the pharmacognosy of Hyptis suaveolens leaf including its morphological, anatomical, chemical constituents and powder analysis. Contrary to earlier reports, the leaves are amphistomatic. Stomata are diacytic, a few being isotricytic and ansiotricytic. Four types of trichomes have been noted. The venation is semicraspidodromous. Midrib consists of one large moon shaped and petiole with four wedge-shaped vascular bundles. Powder microscopy revealed fragments of epidermis, mesophyll, trichomes and tracheary elements. Positive tests for glycosides, polyphenolases, flavones and juglones are also recorded.

INTRODUCTION

The leaves of **Hyptis suaveolens** are considered to be a stimulant, carminative, sudorific and lactogogue. Infusion of the plant is used in catarrhal conditions, affections of the uterus and parasitical cutaneous diseases. The **munda** tribe uses the plant for headache and as snuff to stop bleeding of the nose (1), (2).

The leaves are used as green manure in certain parts of Indian west coast. The shoot tips along with leaves are edible and are sometimes used for flavouring. They are also used as an adulterant for **Patchouli** (**Pogostemon**) leaves. An infusion of leaves is taken as beverage. In Java, the plant is used as cattle fodder (3). No detailed information is available on the morphology, anatomy and pharmacognosy (3 - 6) and hence the present study is taken up.

MATERIALS AND METHODS

Twigs of **Hyptis suaveolens** were collected from plants growing in Osmania University campus, Hyderabad, Andhra Pradesh. The methodology and terms used for after Prabhakar and Ramayya (7), Prabhakar **et al** (8), Leelavathi **et al** (9), Prabhakar and Leelavathi (10).

Abbreviations used in the text are: Ab: abaxial surface of the organ, Ad: adaxial surface of the organ, D: diameter, Dist: distribution, E. C. F. epidermal cell frequency, L|W: length | width, S|F,: Stomatal frequency.

Observations and Discussion

Hyptis is a cosmopolitan genus represented by 400 species (11) of which **H.suaveolens** is reputed for its medicinal uses and is widely distributed having been recorded throughout India, ascending from Himalayas upto 6000 feet and Ceylon, Thwautes, Malaya islands to Australia, West Asia and Arabia (2).

Vernacular names:

BENGAL : Billati tulsi, BIHAR : Bhusri, Dimbubha, Gusumpuru, HINDI : Wilayati tulasi, ORIYA : Ganga tulsi (1), (2), (3).

Morphology:

Annual, erect, branched, hirsute, Habit: glandular - pubescent, aromatic herb, upto 2m high. Root : Tap root, Stem : Green quandrangular, hairy, woody below. Leaf: Simple, opposite, cordate with rounded or subcordate base, pubescent. Margin serrate, apex acute, 7 x 6.5 cm. Petiole: 3.5 cm, long slender, hairy. Inflorescence: axillary cymes, pubescent, few-flowered, peduncle 1.5 cm. Flower: Blue, complete, zygomorphic, pedicillate, pedicels longer than calyx, bracteolate, Bract: subluate, 2mm long, **Bracteoles:** 2, bract-like 1 - 1.5mm. Calyx : some what bilabiate; tube 5 -6 mm; lobes subequal, subulate, 2mm. **Corolla:** Bilabiate, pubescent; tube 9 – 10 mm; lower lip saccate, upper rounded. Androecium: stamens four, exerted. didynamous, inserted below the throat, declinate; filaments and attachement densely villous, longer filaments 3 mm; anther-lobes confluent, 0.75 mm. Ovary: deeply 4 lobed with a single basal ovule in each lobe, 0.5 mm; style gynobasic, 4 mm; stigma bifid. Fruit: aggregate of 4 mules. Flowering and Fruiting: October to December.

Microscopic Characters:

Surface view of leaf: Anticlinal walls of laminar epidermal cells were described to be wavy to sinuate in the subtribe Hyptidinae (13). In the present taxon, the epidermal cells are polygonal anisodiametric and rarely polygonal liner. Aniticlinal walls are sinuate, sinuses being 'U' – and 'V' – shaped. Walls are slightly thick and surface is smooth. Cytoplasmic contents are scanty. Cells are irregularly arranged and variously oriented. **Dist:** All over the leaf lamina except on veins (Plate I. 1,2).

Foliar costal cells and stomatal distribution patterns were considered to he of identification value (9). Unlike in other angiospermous taxa presently it is observed that the costal cells are mostly polygonal anisodiametric and rarely linear in shape, with scanty cytoplasmic contents. Anticlinal walls are straight to curved, and slightly They are parallel oriented and thick. irregularly arranged. **Dist:** Present on all grades of veins.

Type of the stomata in **H. suaveolens** was reported to be diacytic (13), (14), which is confirmed presently besides. their association with isotricytic and anisotricytic stomata and all are monocylic (Plate I. 1,2). Shape of the guard cells are elliptic and flushed with epidermal cells. Size of the stomata on adaxial and abaxial surface is 23 $-20 - 15 \,\mu m$ (length) and $19 - 15 - 11 \,\mu m$ (width). Subsidiaries are mostly of 'a' type rarely of 'f' type, and are indistinct from epidermal cells (Plate I. 1,2). Dist: The leaves were reported to be hypostomatic (14). However presently they are observed to be amphistomatic and distributed all over the leaf lamina except on veins of both the surfaces (Plate I. 1, 2).

In the past, glandular (uni or bicelled head) and non glandular (uniseriate and multicellular) trichomes were reported (13), (15), (16), (17). However presently in **H. suaveolens** four types of trichomes, distributed all over except on margin are observed which are described below: **Uniseriate conical hair: Foot:** one celled; embedded or slightly projected above the epidermis, indistinct, contents scanty; walls thin. **Body:** conial, 2 to 5 – celled in length; cells longer than broad; contents scanty; walls thin; surface smooth (Fig. 4).

Uniseriate capitates hair: Foot: one celled, embedded or slightly projected above the epidermis indistinct; usually broader than long; contents slightly dense; walls thin. Stalk: one – celled; broader than long; contents slightly dense; walls thin; surface smooth. Head: globose, 1-celled, contents dense, walls thin, surface smooth (Fig. 6).

Uniserate pelate hairs: Foot: one-celled, embedded or slightly projected above the epidermis indistinct; broader than long; contents scanty; walls thin. **Stalk:** One – celled broader than long; contents scanty; walls thin surface smooth. **Head:** two or four-celled, cells longer than broad; contents dense; walls thin; surface smooth (Fig. 7).

Uniserate vesicular peltate hairs: Similar to peltate hairs, but the head is enclosed by a thin cuticular vesicle (Fig. 7).

Foliar venation as a taxonomic tool has been since long time in use (7), which is equally important in pharmacognostic studies (18). In **H. suaveolens** the venation was reported to be smi-craspidodromous in which a branch of secondary vein is joining the superdijacent secondaries forming a loop while the others terminating in the margin (Rudal, 1980) which is presently confirmed (Fig. 5). Tertiaries are percurrent and their angle of origin is acute; right angel (AR). The relationship of the tertiaries to midvein are oblique throughout but perpendicular distally and their course is straight to retroflexed and predominantly opposite. Higher order is observed upto 5° ; Areoles are polygonal, regular in orientation and $9|\text{mm}^2$; veinlets simple curved, branched once, 0-1 | areole and 6 | mm².

Sectional view of leaf: In transaction, leaf lamina is ribbed abaxially at primary, secondary, tertiary and alveolar veins but adaxially only at primary vein (Plate I. 3). Epidermal cells are circular to oval shaped, abaxial ones smaller in size (lamina Ad: L|W: 31 - 21 - 15 - | 15 - 12 - 11; Ab;L|W: 19 - 15 - 11 | 15 - 10 - 08; midrib: L|W; $23 - 15 - 11 | 15 - 11 - 8 \mu m$;) with thin cuticle and scanty contents. Outer wall of the guard cells are ledged. Palisade cells are irregular in shape (L | W 57 - 42 - 30 | $11 - 9 - 8 \mu m$) with large intercellular spaces. Spongy mesophyll cells are also irregular in shape (D = $19 - 11 - 8 \mu m$). Near midvein and major lateral veins on both sides, hypodermis is compactly arranged 1 to 3 layered angular collenchyma $(D - 16 - 12 - 11 \mu m)$ and rest of the ground tissue is parenchymatous. Parenchyma 5 to 6 – layered; cells are polygonal in shape $(D = 49 - 38 - 19 \mu m)$ Midvein consists of central large collateral lunar shaped vascular bundle which is apreicyclic (Plate I. 3). Tracheary elements are polygonal to circular (D = 19 - 12 - 8µm) numerous in midvein and secondary veins, few in others, arranged in rows. Secondary wall thickenings annular (free), helical (helices single or double). Perforation plate is simple (Figs. 11 - 13).

In transaction the petiole is semi circular and adaxially groved with lateral ridges (Plate I. 4). Epidermal cells are circular to oval in shape ($D = 15 - 12 - 11 \mu m$). Ground tissue is heterogenous with 3 to 5 – layered compactly arranged collenchymatous hypodermis; cells being circular to polygonal ($D = 19 - 17 - 15 \mu m$) with scanty contents. Rest of the ground tissue is

5 to 8 layered parenchymatous. Cells are polygonal (D = $68 - 43 - 30 \mu m$) with thin walls and scanty contents. Vascular bundles are four wedge shaped. Adaxial two bundles are larger compared to the two abaxial ones. Tracheary elements are polygonal, few circular (D = $23 - 16 - 11 \mu m$) arranged in rows. Secondary wall thickenings are annular (free), helical (helices single or double) with simple perforations.

Powder study

The powder is brown aromatic. Microscopic examination of the powder revealed the following elements. 1) Fragments of epidermis with sinuate anticlinal walls and diacytic stomata (Fig. 1, 2); 2) Fragments of castae or petiole showing small polygonal epidermal cells without stomata (Fig. 3); 3) Full or bits of uniseriate conical, capitates, peltate and vesicular peltate hairs (Fig. 4 - 7); 4) Groups or isolated palisade, spongy and ground parenchyma and collenchymas (Fig 8 - 10). 6) Bits of vessels with annular and helical thickenings (Figs. 12 – 13).



34



Figs. 1-3, 5, 8-13; Hyptis succeedens: L. leaf powder showing: 1 & 2; Pieces of epidermis showing epidermal cells & stomata; 3. Costal cells; 5. Full or bits of trichomes; Figs. 4, 6, 7; Trichomes in sectional views: 4. Uniseriate conical hair; 6. Uniseriate capitate hair; 7. Uniseriate vesicular peltate hair; 8. Palisade parenchyma; 9. Spongy parenchyma; 10. Collenchyma cells; 11. Ground parenchyma cells; 12 & 13. Tracheary elements.

Besides the microscopic observations the cold extracts of the powder in different solvents and the moist and dry powders are observed under ordinary and ultraviolet light to record the colour changes (colours were compared with colour index (19) and details are given in table 1.

Following the simple phytochemical test recommended by Gibbs, (20) for identification of chemical constituents in the fresh dried leaf powder. Positive tests were recorded for glycosides, polyphenolases, flavones, juglones and negative tests for saponins, syrengins, aucubins, aurones, and leucoanthocynins, which will be much helpful in identification of the taxa in powder form.

In the light of the present investigation the salient features of the drug which would be

useful in identification in its different states are as follows:

Lamina: Leaves are ovate, base subcordate apex acute; margin serrate hairy, dark green 7 x 6.5 cm with unicostate reticulate semicraspidodromous venation. Secondary veins 9 - 10 pairs. Epidermal cells mostly polygonal anisodimetric sides are sinuate on both adaxial and abaxial. Costal cells distinct on all veins. Leaves anphistomatic, stomata mostly diacytic, with a few isotricytic and anisotricytic, monocyclic. Uniseriate conial, capitates, peltate and vesicular peltate hairs present all over except on margins. In the transaction, leaf is bifacial, with one layered palisade and 5 - 6layers of spongy mesophyll. Midvein with single large lunar shaped vascular bundle.

TABLE 1

Sl. No.	Powders	Ordinary Light	Ultraviolet Light
1	Dry Powder	BB*	DP
2	Aquous extract	GB	DB
3	Aquous residue	DB	DB
4	Alcoholic extract	DB	BB
5	Alcoholic residue	BB	DB
6	Chloroform extract	DB	DB
7	Chloroform residue	BB	GP

Observations of leaf powder under ordinary and U. V. Light

**BB. Brown black; DB: Dark brown; DP: Dull brown; GB: Gold brown; GP: Greenish brown.

Petiole: 3.5 cm long, slender, densely hairy. In transaction it is semicircular and adaxially grooved with lateral ridges. Ground tissue is heterogenous consisting of 3 - 5 layered compactly arranged collenchymatous hypodermis and 5 to 8 – layered parenchymatous tissue. Vascular bundles are 4 wedge shaped.

Leaf powder: Leaf powder is brown and aromation: Microscopically fragments of

epidermis, stomata, mesophll, trichomes and trachary elements can be observed. Positivies tests for glycosides, polyphenolases, flavones, juglones are recorded.

Acknowledgements

The authors are thankful to Prof. N. R. Subb Raju, Head Department of Botany, Osmania University, Hyderabad, A. P. for the facilities and encouragement.

REFERENCES

- 1. Kirthikar, K. R. Basu, B. D. Indian Medicinal Plants, Vol.II, Allahabad (1933).
- 2. Nadkarni, A. K. Dr. K. M. Nadkarni's Indian Materia Medica, Bombay, Popular Prakashan (1976).
- 3. Chopra, R. N. Nayar, S. L. Chopra, I. C. Glossary of Indian Medicinal Plants, New Delhi, CSIR (1956).
- 4. Youngken, H. K. Textbook of Pharmacognosy, Torento, Blakiston comm. (1950).
- 5. Anonymous. The Wealth of India, Vol. III N-Pc, New Delhi, CSIR (1966).
- 6. Anonymous. Homoepathic Pharmacopoea of India, Vol. I, Govt. of India, New Delhi, Ministry of Health (1971).
- 7. Prabhakar, M. Ramayya, N. Foliarvnation patterns and their taxonomic importance in Indian Portulaceae. Geophytology (12(1)): 49 – 54 (1982).
- 8. Prabhakar, M. Ramayya, N. Leelavathi, P. Structure and distribution of epidermal elements in the angisoperms 1. Epidermal cells comple. Geophytology 14: 55 – 68 (1984).
- 9. Leelavathi, A. Aamayya, N. Nrabhakar, M. Study of the leaf costal distribution patterns and their significance in Leguminosae. Geophytology 11: 125 – 135 (1981).
- 10. Prabhakar, M. Leelavathi, P. Structure, delimitation nomenclature and classification of plant trichomes. Asia Jour. Pl. Sci. 1: 49 – 66 (1989).
- 11. Airy Shaw, H. K. Willis A Dictionary of the Flowering Plants and Ferns, 7th ed. Cambridge, University Press (1966).
- 12. Hooker, J. D. Flora of British India, Vol. II, Lond. Reeve and Co., Ltd. (1879).
- 13. Rudal, P. Leaf anatomy of the subtribe Hyptidinae of Labiatae. Bot. Jour. Linn. Soc. 80 : 319 - 340 (1980).
- 14. Inamdar, J. Bhatt, D. C. Structure and stomatal ontogeny in some Labiatae. Ann. Bot. 36: 325 – 334 (1972).
- 15. Sclereder, H. Syste, atoc Anatomy of Dicotyledons, Vol. I, Oxford, Clarenden Press (1908).
- 16. Korsmo, E. Anatomy of weeds, Oslo, New York, Momliv (1954).

- 17. Metcalfe, C. R. Chalk, L. Anatomy of Dicotyledons, Vol. II, Clarendon Press, Oxford (1950).
- 18. Leelavathi, P., Prabhakar, M. Ramayya N. Pharmacognostic studies on the leaf of Acalypha medica L. Indian J. Bot. 11: 129 138 (1988).
- 19. Maerz, A. Paul, M. A. Dictionary of Colour (II ed.) McGraw-Hill Book Co. Inc. London (1950).
- 20. Gibbs, D. Chemotaxonomy of Flowering Plants, Vol. I, II and III Mc Gillqueens University, Montreal and London (1974).